

# ImmunoComb®

BOVINE *CHLAMYDOPHILA SP.*  
(*CHLAMYDIA SP.*)  
ANTIBODY TEST KIT

## INSTRUCTION MANUAL SUFFICIENT FOR 30 ASSAYS

### I. INTENDED USE

This kit is designed to determine cow serum IgG antibody titers for *Chlamydomphila sp.* (previously known as *Chlamydia sp.*).

### II. WHAT IS THE ImmunoComb® ASSAY?

The ImmunoComb® is a self-contained portable kit. A sensitive test which detects antibody levels in the blood or serum, the ImmunoComb® provides results within 35 minutes.

### III. HOW DOES THE ImmunoComb® WORK?

- Based on a solid phase immunoassay principle, the ImmunoComb® is a plastic card shaped like a comb, on which purified Chlamydomphila antigen is attached.
- Either immerse paper disks in cow blood or take a serum specimen. Deposit into sample wells of the multi-compartment developing plate.
- Insert Comb into the sample wells so that antibodies from samples bind themselves to the antigens on the Comb.
- Non bound antibodies are washed out in the second compartment.
- The next compartment contains an anti-cow IgG antibody labeled with an enzyme.
- Immerse the Comb in this "conjugate." The bound antibodies will be labeled.
- Insert the Comb into a compartment where the enzyme reaction takes place. This generates a color change which indicates the amount of antibodies present.
- Using the CombScale, convert the spot's color intensity to the anti-chlamydomphila immunoglobulin level.
- An internal control, the top spot indicates that the development is completed.
- The ImmunoComb® may be divided into three separate sections. Each segment processes between 1-4 samples.

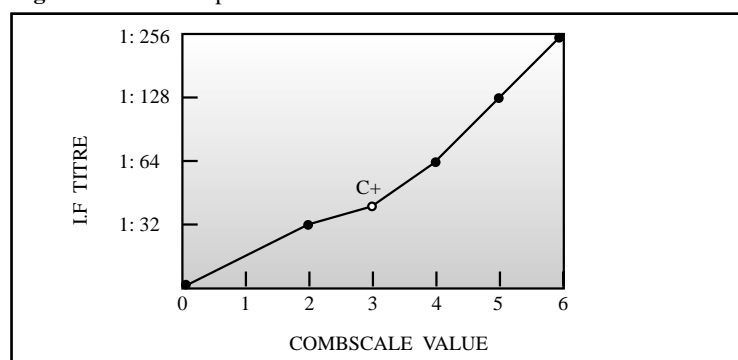
### IV. HANDLING & STORAGE

1. Store the kit under normal refrigeration: 2° - 8° C (36° - 46° F).
- Do not freeze the kit.**
2. Before conducting the test, maintain all kit elements and specimens at room temperature -- preferably for 60-120 minutes (or 22 minutes in 37° C). Perform assay at room temperature of 20° - 25° C (68° - 77° F).
  3. Avoid spillage and cross-contamination of solutions.
  4. Mix reagents by inverting developing plate several times prior to use.
  5. Do not mix reagents from different kits or from different compartments of one kit.
  6. Do not touch teeth of ImmunoComb® Card.
  7. When using developing plate, pierce cover of each compartment while strictly following test procedure instructions. **DO NOT RIP OFF OR REMOVE COVER OF ENTIRE DEVELOPING PLATE ALL AT ONCE.**
  8. The ImmunoComb® kit contains inactivated biological material. Kit must be handled and disposed of in accordance with accepted sanitary requirements. It is recommended to incinerate kit after use. Use large amounts of water to flush kit solutions down sewage/drainage system.

### V. READING AND INTERPRETING THE RESULTS

- To determine the IgG titer of Chlamydomphila specimens, compare the color intensity of the Comb's appropriate teeth with the color spot series on the enclosed CombScale table (see illustrations 9 & 10 for details).
- The bottom spot on the ImmunoComb® tests for Chlamydomphila. Evaluate the results of each spot separately.
- Compare the specimen's color intensity with that of the positive control (C+) included in the kit, in order to determine its titer.
- The positive control (C+) is calibrated to a 1:32 titer I.F.
- Specimens with an identical or higher color intensity than the positive control are considered positive.
- The negative control consists of non-immuned sera and should be read as zero (S=0).
- Specimens with a color intensity lower than the positive control are considered negative or non-immuned.
- To evaluate the titer, use the CombScale provided in the kit and determine the titer using Fig. A, as reference. The procedure is detailed in section VI.

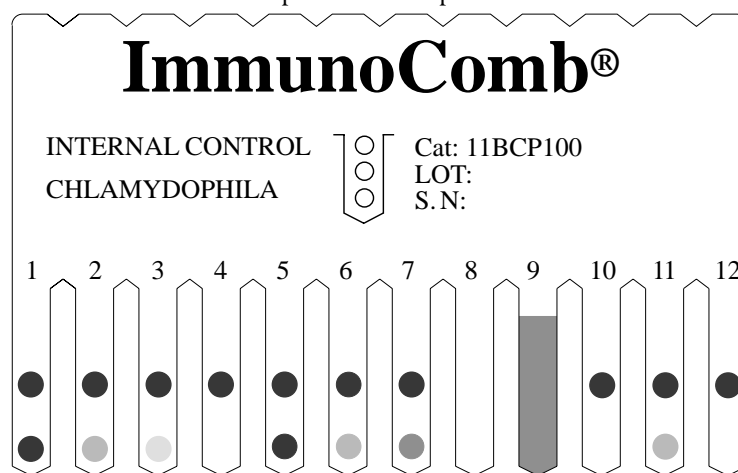
**Fig. A.** Relationship between the CombScale's "S" value and the I.F.



**Important**

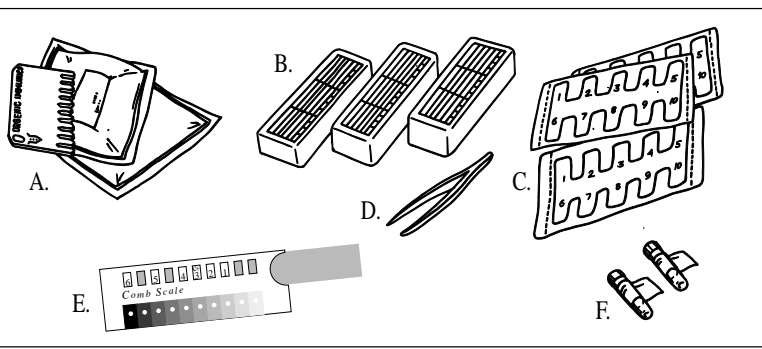
The margin of error is similar to that of other enzyme immunoassay kit procedures. Therefore, an error in one color scale window will not result in a wrong diagnosis.

Example of a developed Comb



TOOTH No.	RESULT & REMARKS
1,5	High positive reaction for Chlamydomphila.
2,6	Positive reaction for Chlamydomphila.
3	Very Low reaction for Chlamydomphila - Considered negative.
4, 10	Negative reaction for Chlamydomphila.
7	Medium positive reaction for Chlamydomphila.
8	No internal control - development failed.
9	High back ground color - no valid test.
11	Positive control.
12	Negative control.

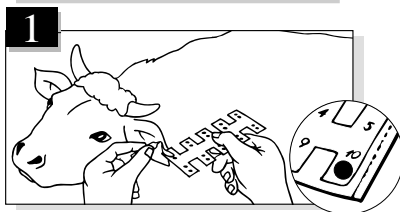
# STEP-BY-STEP WITH ImmunoComb®



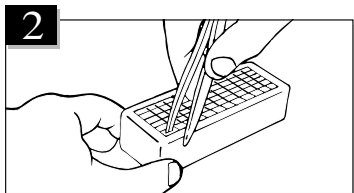
The ImmunoComb® kit includes: A. 3 ImmunoComb® cards, each separately wrapped in an aluminum envelope; B. 3 developing plates; C. 3 specimen papers with pre-punched disks; D. One disposable tweezers; E. One calibrated CombScale color card; F. One tube of positive control serum and one tube of negative control serum, a CombScore sheet and a user manual.

Perform assay at room temperature of 20° - 25° C (68° - 77° F).

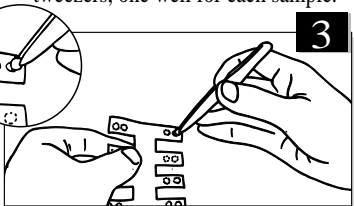
## When using a paper disk



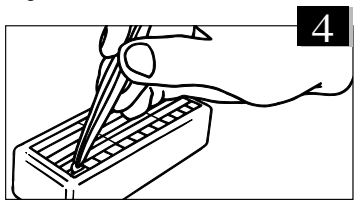
Pierce one of the cow's veins. Take a specimen paper and saturate a pre-punched disk with the blood.



Slit open the protective aluminum covering of compartment A with the tweezers, one well for each sample.



When using a paper disk punch out a disk saturated with blood.



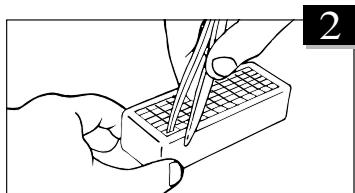
Insert the disk into well #1 of compartment A. Dip it well into the liquid. Proceed with the other samples.

Wait 60 minutes for extraction of antibodies.

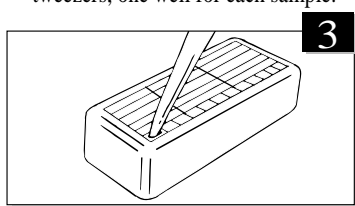
## When using a serum



Use a pipette or a capillary tube. For testing a serum sample use 5 µl.



Slit open the protective aluminum cover of compartment A with the tweezers, one well for each sample.



Dispense a sample into each well. When using the capillary tubes raise and lower the piston several times to achieve mixing. When using a pipette, mix by depressing the plunger a number of times.

Proceed to step 5.

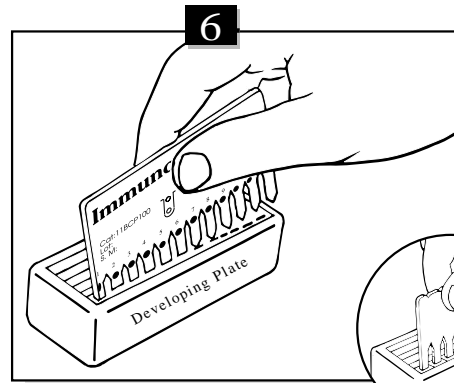
**5**

For control serum open the next 2 consecutive wells.

Take 5 µl positive control serum (C+) and insert into well A next to the last sample.

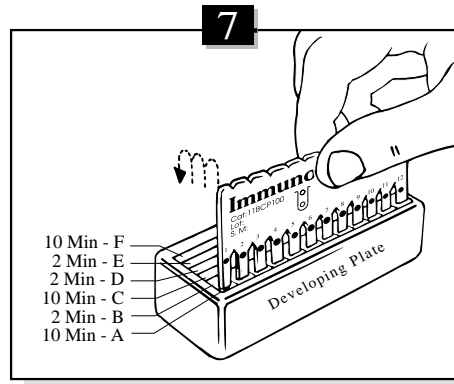
Mix the serum in the well

Do the same with the negative control serum in the next well.

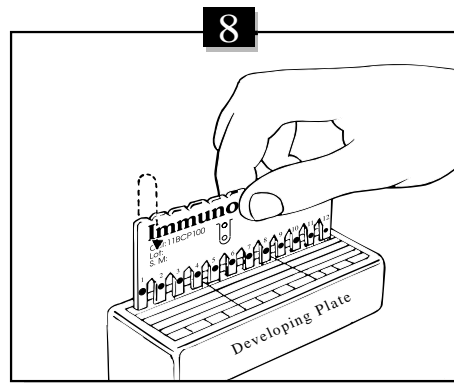


Remove one ImmunoComb® from its protective wrapping and insert (print side facing you) in compartments of **Row A**. Gently move Comb up and down several times, then let incubate in **Row A's** compartments for **10 minutes**.

When using 1/3 or 2/3 of a Comb, break the Comb by folding back on notch 4 or 9 respectively. Keep the rest in its original sleeve for further use.



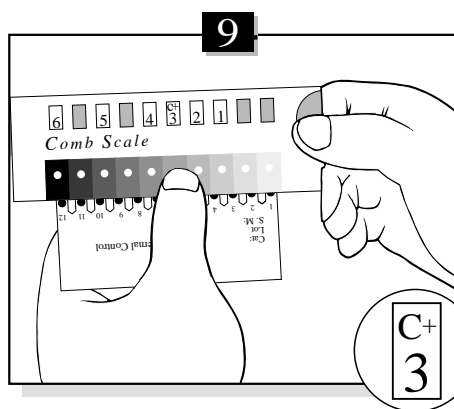
Pierce the cover of the appropriate section of compartment **B** with the tweezers. Follow same procedure for remaining rows at end of each incubation period. Gently shake off excess liquid onto a tissue. Insert Comb in **row B's** compartment and let incubate for **2 minutes**, shake-off and transfer Comb to **Row C** and incubate for **10 minutes**. Similarly, the Comb is placed in **Row D** for **2 minutes**, **Row E** for **2 minutes**, and **Row F** for **10 minutes**, allowing the color reaction process to develop.



After the Comb has completed the cycle for **Row F**, transfer it back to **Row E**. Incubate in **Row E** for **2 minutes** to fix color.

**AIR DRY  
AND READ  
RESULTS**

## VI. READING RESULTS WITH THE CombScale



### A. Adjust scale with positive control:

When the Comb is completely dry align it with the calibrated color CombScale. Compare the color resulting from the positive control (C+) sample to the color scale: slide the yellow ruler until the "C+" mark appears in the window corresponding to the color.

FINALLY, HOLD THE SLIDE IN THIS POSITION DURING READING.

### B. Read each of the spots separately:

Choose the most suitable color and read the titer in the yellow windows.

**REMEMBER: A DIFFERENCE OF ONE COLOR LEVEL WILL NOT AFFECT DIAGNOSIS !!!**

